

## APOTRICHOTHECENE REARRANGEMENT IN MACROCYCLIC TRICHOTHECENE DERIVATIVES [1]

N. Jeker and Ch. Tamm \*

Institute of Organic Chemistry, University of Basel,  
St. Johannis-Ring 19, CH-4056 Basel, Switzerland

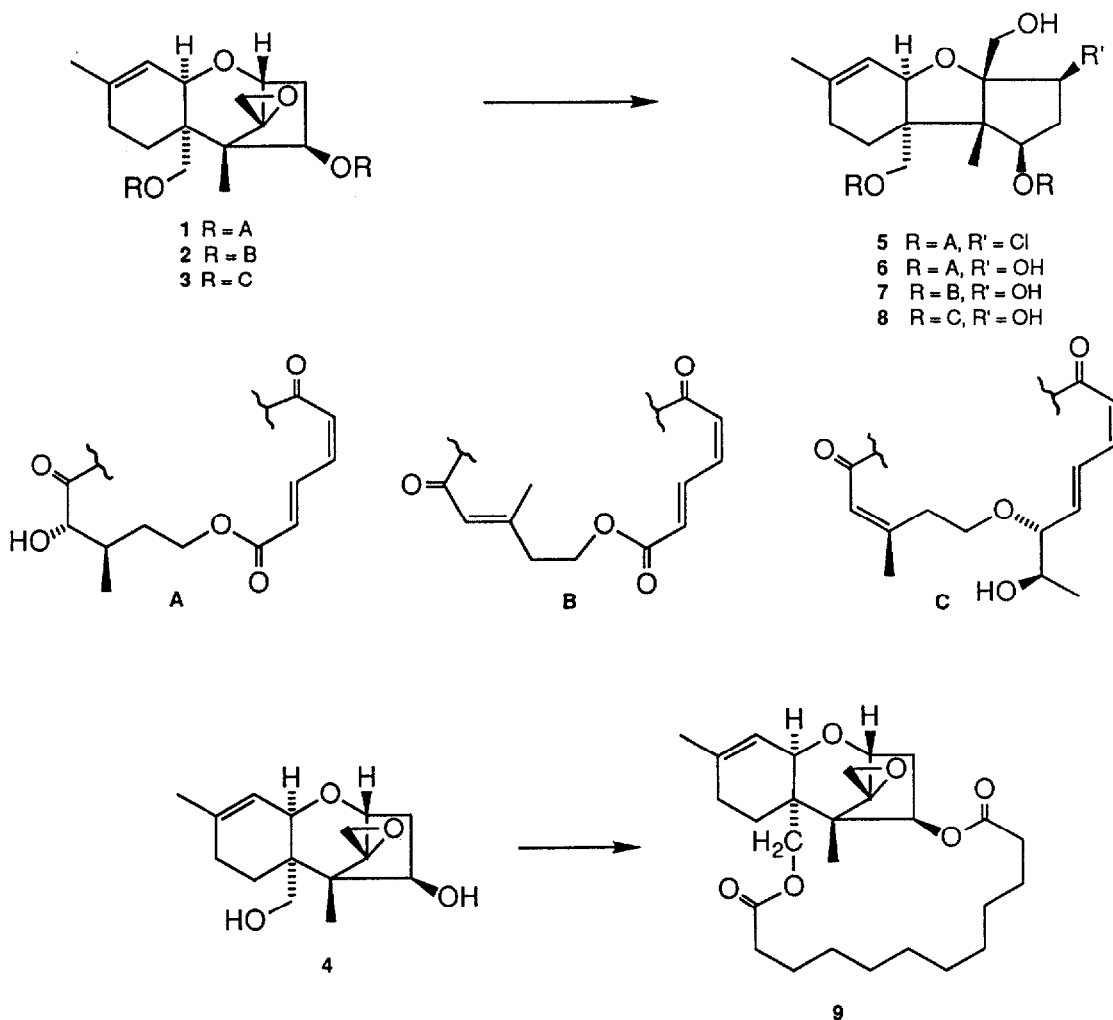
**ABSTRACT:** The apotrichothecene rearrangement was carried out with verrucarin A (1), J (2) and roridin A (3). The synthesis of the macrocyclic diester 9 of verrucarol (4) is described. The immunosuppressive (MLR) and cytotoxic (P-815) activities of the compounds 6-9 were determined *in vitro*.

The trichothecenes belong to a class of sesquiterpenoid natural products which are produced by moulds, especially various species of *Fungi imperfecti*. Many members of the family display a wide range of biological activities [3]. The rearrangement of the trichothecene skeleton during treatment with mineral acids is well known and leads by initial cleavage of the oxirane ring to the apotrichothecene skeleton which is biological inactive [4,5,6]. In connection with our programme directed towards the synthesis of unnatural macrocyclic trichothecene derivatives [7] we investigated the apotrichothecene rearrangement of the macrocyclic trichothecenes verrucarin A (1), J (2) and roridin A (3). Our interest was focused on the question whether this rearrangement generally takes place without touching the macrocyclic moiety of the mycotoxins and on the biological activity of such rearranged macrocyclic products which to our knowledge had not yet been examined.

The trichothecenes 1, 2 and 3 were treated with H<sub>2</sub>SO<sub>4</sub> in dioxane. In contrast to the simple trichothecene verrucarol (4) [4] the rearrangement of the macrocyclic derivatives proceeded at higher temperature (80°, 4-6h) and led to the products 6, 7 and 8 with intact macrocyclic segments. The analogous rearrangement of 1 with HCl in CH<sub>2</sub>Cl<sub>2</sub> was reported by Grove [6] and led to the chloropotrichothecene 5. The structures of the products obtained were deduced from the MS and NMR data.

In order to examine the relations between the substitution pattern of the macrocyclic segment and the biological activity, the simple 18-membered macrocyclic trichothecene 9 was synthesized using dodecanedioic acid as building block for the macrocyclic part. Verrucarol (4) was esterified with the mixed dianhydride of dodecanedioic acid and pivalic acid leading to the macrocyclic diester 9 in about 30% yield.

**Biological Activity:** The cytotoxic and immunosuppressive activities of the new compounds were determined by *P.Hiestand, Sandoz AG Basel*, using the P-815 [8] and MLR [9] *in vitro* tests. The rearranged compounds **6 - 8** showed a weak immunosuppressive (MLR) activity (  $IC_{50}$  values in  $\mu\text{g/ml}$ : **6**: 0.07, **7**: 0.05, **8**: 0.05 ) On the other hand **6 - 8** were found to be inactive in the cytotoxicity (P-815) test. (  $IC_{50}$  values for **6 - 8**:  $> 1\mu\text{g/ml}$ ). The macrocyclic analogue **9** was found to be moderately active (MLR: 0.2  $\mu\text{g/ml}$ , P-815: 0.088  $\mu\text{g/ml}$ ). It's activity was shown to be about hundred times lower than that of natural macrocyclic compounds, a fact which demonstrates the necessity of a suitably functionalized macrocyclic segment to obtain a high biologically active compound.



Experimental Procedures: Apotrichothecene Rearrangement: A soln. of 54 mg of verrucaric acid (1) in 2.5 ml of dioxane and 1.5 ml of 1N H<sub>2</sub>SO<sub>4</sub> was heated to 80°. After 6h the dioxane was distilled off and the aqueous layer extracted with CH<sub>2</sub>Cl<sub>2</sub>. Column chromatography (SiO<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub> / MeOH 96:4) afforded 23 mg (41%) of 6. (The same procedure yielded 59% of 7 and 73% of 8, respectively.)

Characterization of 6: m.p.: 230-300°: dec. (from CH<sub>2</sub>Cl<sub>2</sub> / hexane).  $[\alpha]_D^{23} = +117^\circ$ . EI-MS: 520 (M<sup>+</sup>, 10.0%), 265 (6.2%), 248 (4.7%), 189 (7.2%), 175 (12.7%), 163 (43.0%), 145 (13.5%), 125 (21.1%), 105 (100%), 85 (57.1%), 55 (31.6%). <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>, Me<sub>4</sub>Si): 8.46 (*ddd*, *J* = 1, 11.5, 15.5, H-C(4'')); 6.77 (*dd*, *J* = 11.0, 11.5, H-C(3'')); 6.10 (*dd*, *J* = 1, 11.0, H-C(2'')); 6.06 (*d*, *J* = 15.5, H-C(5'')); 5.91 (*d*, *J* = 6.5, H-C(4)); 5.51 (*ddd*, *J* = 1.5, 1.5, 5.0, H-C(10)); 4.57 (*m*, H-C(5'')); 4.56 (*d*, *J* = 12.5, H-C(15)); 4.43 (*dd*, *J* = 2.5, 6.0, with D<sub>2</sub>O: *d*, *J* = 2.5, H-C(2'')); 4.28 (*dd*, *J* = 5.0, 5.0, with D<sub>2</sub>O: *d*, *J* = 5); 4.20 (*br.d*, *J* = 12.5, H-C(15)); 4.05 (*dt*, *J* = 3.0, 11.5, H-C(5'')); 3.96 (*dd*, *J* = 8.5, 12.0, with D<sub>2</sub>O: *d*, *J* = 12.0, H-C(13)); 3.80 (*dd*, *J* = 4.5, 12.0, with D<sub>2</sub>O: *d*, *J* = 12.0, H-C(13)); 3.67 (*d*, *J* = 5.0, H-C(11)); 2.61 (*d*, *J* = 6.0, exchangeable with D<sub>2</sub>O, OH); 2.50 (*m*, H-C(3'')); 2.35 (*d*, *J* = 6.0, exchangeable with D<sub>2</sub>O, OH); 2.21 (*m*, exchangeable with D<sub>2</sub>O, OH); 2.19 (*ddd*, *J* = 5.0, 6.5, 15.5, H-C(3)); 2.05 (*d*, *J* = 15.5, H-C(3)); ca. 1.93 (*m*, H-C(4'')); ca. 1.75 (*m*, H-C(4'')); 1.74 (*br.s*, H<sub>3</sub>C(16)); 1.03 (*s*, H<sub>3</sub>C(14)); 0.90 (*d*, *J* = 7.0, H<sub>3</sub>C(6')). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>, Me<sub>4</sub>Si): 174.9, 165.4, 164.8, 140.4, 140.2, 139.4, 127.4, 124.8, 118.0, 96.1, 80.1, 79.4, 75.3, 73.6, 64.6, 63.4, 61.9, 59.9, 47.9, 39.6, 33.2, 32.2, 27.5, 23.3, 22.7, 13.0, 11.9.

Characterization of 7: m.p. 242- >300°: dec. (from CH<sub>2</sub>Cl<sub>2</sub> / hexane).  $[\alpha]_D^{23} = -26^\circ$  EI-MS: 502 (M<sup>+</sup>, 5.8%), 265 (5.5%), 248 (6.3%), 217 (16.5%), 189 (6.8%), 175 (13.1%), 163 (43.4%), 145 (16.3%), 125 (23.6%), 113 (100%), 95 (99.0%), 80 (54.8%), 67 (56.0%), 52 (59.5%). <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>, Me<sub>4</sub>Si): 8.40 (*ddd*, *J* = 1, 11.5, 16.0, H-C(4'')); 6.68 (*ddd*, *J* = 1, 11.5, 11.5, H-C(3'')); 6.03 (*dd*, *J* = 1, 11.0, H-C(2'')); 6.00 (*d*, *J* = 16.0, H-C(5'')); 5.85 (*m*, H-C(4)); 5.83 (*s*, with D<sub>2</sub>O: *d*, *J* = 1, H-C(2'')); 5.52 (*m*, H-C(10)); 4.56 (*ddd*, *J* = 5.0, 5.0, 11.5, H-C(5'')); 4.48 (*d*, *J* = 12.5, H-C(15)); 4.27 (*m*, with D<sub>2</sub>O: *dd*, *J* = 3.0, 5.0, H-C(2)); 4.09 (*ddd*, *J* = 6.5, 6.5, 11.5, H-C(5'')); 3.94 (*dd*, *J* = 9.0, 11.5 with D<sub>2</sub>O: *d*, *J* = 11.5, H-C(13)); 3.84 (*dd*, *J* = 4.0, 11.5, with D<sub>2</sub>O: *d*, *J* = 11.5, H-C(13)); ca. 3.82 (H-C(11)); 3.77 (*d*, *J* = 12.5, H-C(15)); ca. 2.52 (exchangeable with D<sub>2</sub>O, OH); ca. 2.32 (exchangeable with D<sub>2</sub>O, OH); 2.25 (*d*, *J* = 1.5, H<sub>3</sub>C(6')); 1.72 (*br.s*, H<sub>3</sub>C(16)); 1.06 (*s*, H<sub>3</sub>C(14)).

Characterization of 8: m.p. 175-178° (from CH<sub>2</sub>Cl<sub>2</sub> / ether).  $[\alpha]_D^{23} = +78^\circ$ . EI-MS: 550 (M<sup>+</sup>, 0.7%), 249 (21.0%), 212 (43.0%), 163 (17.2%), 131 (12.1%), 105 (48.1%), 85 (100%). CI-MS: 568 ([M+NH<sub>4</sub>]<sup>+</sup>) (46.4%), 284 (100%). FAB-MS: 551 ([M+H]<sup>+</sup>, 14.0%), 154 (100%). <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>, Me<sub>4</sub>Si): 7.78 (*dd*, *J* = 11.5, 15.5, H-C(4'')); 6.69 (*dd*, *J* = 11.5, 11.5, H-C(3'')); 6.04 (*dd*, *J* = 3.5, 15.5, H-C(5'')); 5.73 (*d*, *J* = 11.0, H-C(2'')); 5.84 (*dd*, *J* = 2.0, 6.0, H-C(4)); 5.51 (*m*, H-C(10)); 4.39 (*d*, *J* = 12.5, H-C(15)); 4.23 (*m*, with D<sub>2</sub>O: *dd*, *J* = 2.5, 5.0, H-C(2)); 4.16 (*d*, *J* = 12.5, H-C(15)); 3.95 (*br.d*, *J* = 11.5: with D<sub>2</sub>O sharp *d*, *J* = 12.0)); 3.81 (*br.d*, *J* = 11.5, with D<sub>2</sub>O: sharp *d*, *J* = 12.0, H-C(13)); 3.72 (*br.d*, *J* = 5.0, H-C(11)); ca. 3.64 (*m*, H-C(6'')); ca. 3.62 (*m*, H-C(7'')); 3.55 (*m*, H<sub>2</sub>C(15)); 2.77 (*br.s*, exchangeable with D<sub>2</sub>O, OH); 2.68 (*d*, *J* = 6.0, exchangeable with D<sub>2</sub>O, OH); 2.52 (*br.s*, exchangeable with D<sub>2</sub>O, OH); ca. 2.30 (*br.s*, exchangeable with D<sub>2</sub>O, OH); ca. 2.28 (*m*, H-C(3'')); ca. 2.27 (*m*, H-C(3)); ca. 1.98 (*m*, H-C(3)); 1.73 (*br.s*, H<sub>3</sub>C(16)); 1.19 (*d*, *J* = 6.0, H<sub>3</sub>C(8'')); 1.08 (*s*, H<sub>3</sub>C(14)); 1.02 (*d*, *J* = 6.5, H<sub>3</sub>C(6')). <sup>13</sup>C-NMR (400 MHz, CDCl<sub>3</sub>,

Me<sub>4</sub>Si): 174.6, 164.9, 144.6, 140.2, 139.9, 127.3, 118.1, 117.2, 95.3, 84.4, 79.4, 77.8, 75.1, 74.4, 70.7, 69.3, 64.5, 63.5, 59.4, 47.5, 39.2, 35.2, 34.1, 27.5, 23.3, 22.5, 18.2, 14.4, 13.0.

Preparation of **9**: To a soln. of 149  $\mu$ l of Et<sub>3</sub>N, 49 mg of dodecanedioic acid and 47  $\mu$ l of pivalic acid in 10 ml of CH<sub>2</sub>Cl<sub>2</sub>, 50 mg of dimethylaminopyridine and 70 ml of CH<sub>2</sub>Cl<sub>2</sub> were added. The soln. was stirred for 30 min., then a soln. of 47 mg of verrucarol (**4**) in 1 ml of CH<sub>2</sub>Cl<sub>2</sub> was added dropwise and stirred for 20h. The solvent was evaporated and the residue purified by column chromatography (SiO<sub>2</sub>, ether / petroleum ether (7:3) which yielded 29 mg (30%) of **9**. M.p. 122.4-124.2° (from hexane). EI-MS: 460 (M<sup>+</sup>, 11.9%), 417 (3.4%), 247 (22.0%), 230 (23.6%), 217 (36.4%), 202 (9.8%), 187 (10.0%), 173 (11.8%), 159 (12.2%), 145 (8.7%), 121 (30.1%), 105 (100%), 91 (31.0%), 81 (49.9%), 69 (28.9%), 55 (66.8%). <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>, Me<sub>4</sub>Si): 5.55 (*dd*, *J* = 4.5, 8.0, H-C(4)); 5.42 (*br.d*, *J* = 5.0, H-C(10)); 4.10 (*d*, *J* = 12.5, H-C(15)); 4.06 (*d*, *J* = 12.5, H-C(15)); 3.82 (*d*, *J* = 5.0, H-C(2)); 3.64 (*br.d*, *J* = 5, H-C(11)); 3.14 (*d*, *J* = 4.0, H-C(13)); 2.84 (*d*, *J* = 4.0, H-C(13)); 2.56 (*dd*, *J* = 7.5, 15.5, H-C(3)); ca. 1.98 (*m*, H-C(3)); 1.71 (*br.s*, H<sub>3</sub>C(16)); 0.88 (*s*, H<sub>3</sub>C(14)). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>, Me<sub>4</sub>Si): 173.5, 173.4, 141.1, 118.3, 79.1, 75.3, 67.0, 65.3, 63.2, 48.7, 47.9, 43.4, 37.1, 34.4, 34.3, 28.1, 28.0, 27.7, 27.2, 27.0, 27.0, 26.9, 24.6, 24.5, 23.2, 20.8, 7.6.

## REFERENCES

- [1] 49th Communication on Verrucarins and Roridins [2]
- [2] 48th Communication: Ch.Tamm & N.Jeker, *Tetrahedron* **1989**, *45*, 2385.
- [3] a) Ch. Tamm, *Fortschr. Chem. Org. Naturst.* **1974**, *31*, 63 ;  
 b) J.R. Bamberg and F.M.Strong in "Microbial Toxins"; S.Kadis, A.Ciegler, C.J. Ail,Eds.; Academic Press: New York 1980.  
 c) T.W. Doyle and W.T. Bradner in "Anticancer Agents Based on Natural Product Models"; I.M. Cassidy and J. Duros, Eds.; Academic Press: New York 1980.  
 d) Y. Ueno in "Developments in Food Science 4: Trichothecenes - Chemical, Biological and Toxicological Aspects", Elsevier: Amsterdam - Oxford - London, 1983.
- [4] J.Gutzwiller, R.Mauli, H.P.Sigg & Ch.Tamm, *Helv. Chim. Acta* **1964**, *47*, 2234.
- [5] H.P. Sigg, R. Mauli, E. Fluri & D. Hauser, *Helv. Chim. Acta* **1965**, *48*, 962.
- [6] J.F. Grove, *J. Chem. Soc. Perkin Trans. I*, **1986**, 647.
- [7] a) N. Jeker & Ch.Tamm, *Helv. Chim. Acta* **1988**, *71*, 1895.  
 b) *ibid.*, *Helv. Chim. Acta* **1988**, *71*, 1904.
- [8] H. Stähelin, *Med. exp.* **1962**, *7*, 92
- [9] T. Meo, The MLR in the mouse. in: " Immunological Methods", Eds. L.Lefkovits & B. Pernis, Academic Press: New York, 1979.

(Received in Germany 10 August 1989)